



# Veterinary sample packaging guide

1.

Ensure labelling is readable and indelible.

Clean jars of any external contamination.



2.

Check lids are tightly sealed.

Tape around the lids of jars to reduce leaks. Electrical tape or paraffin tape give a better seal than sticky tape.



3.

Remove all needles, scalpels and gloves.

Submit fluid samples in sterile jars or in plain (clotted) blood tubes, not in syringes or gloves.



4.

Submit blood tubes in foam racks wrapped in plastic film or held together by elastic bands to reduce the chance of breakage in transit. Wrap single blood tubes in absorbent padding.



5.

Place slides of blood films and smears in a slide transport box and seal these in a ziplock bag to prevent formalin fume contamination during transport. (Formalin induces artefacts in smears.)



6.

Use absorbent material (paper, cotton wool) to line the esky in case of leaks. Consider sealing samples in large ziplock bags to reduce the risk of leaks. Note: rigid containers such as eskies (not postbags) are required to prevent breakage of sample tubes.



7.

Place all samples in the esky.

Place cold bricks around the blood, swabs and fresh samples. Fixed samples do not need chilling (but are not harmed by it).



8.

Seal the esky with tape. Place laboratory submission forms in an envelope or ziplock bag and tape to the side of the esky.



9.

Place a courier label on the top of the esky and call courier.



If you suspect an **exotic, reportable or zoonotic disease**, contact DAFWA Diagnostic Laboratory Services (DDLS) sample receipt or the duty pathologist on +61 (0)8 9368 3351 or [DDLS@agric.wa.gov.au](mailto:DDLS@agric.wa.gov.au).

Always use personal protective equipment including gloves when handling biological samples and practice good hygiene.

Current as of November 2016. Check [agric.wa.gov.au](http://agric.wa.gov.au) for updates or contact DDLS – Animal Pathology.